



## Pacific Pests, Pathogens & Weeds - Fact Sheets

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### Taro corm rots - post-harvest (179)



Photo 1. The dry white crumbly rot at the bottom of the corm is caused by *Pythium splendens*. The pinkish tissues above and slightly to the right are a reaction to infection by the corm tissues. The white semi-circle on the left is the cottony growth of *Athelia rolfsii*.



Photo 2. Light brown firm rot caused by *Phytophthora colocasiae*. The rot on the lower left side is caused by *Athelia rolfsii*. The corm has been incubated at high humidity overnight to stimulate the growth of the fungus.



Photo 3. Light brown firm rot in a taro corm caused by *Phytophthora colocasiae*, after about 8 days.



Photo 4. Black spongy rot caused by *Lasiodiplodia theobromae*. These rots usually follow those of *Pythium* and *Phytophthora*, or they come in later after the corms have started to lose moisture.

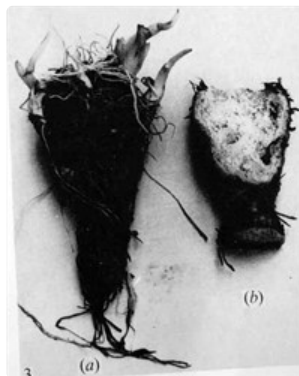


Photo 5. These two corms have been in a plastic bag for 4 weeks; the one on the left is still healthy whereas the one on the right has been destroyed by the soft rot bacterium, *Erwinia chrysanthemi*.

**Common Name**

Taro corm rots (post-harvest)

## Scientific Name

The following are the commonly recorded rots that develop in taro corms after harvest:

*Athelia rolfsii* (see **Fact Sheet no. 11**)

*Lasiodiplodia (Botryodiplodia) theobromae*

*Pythium splendens*

*Phytophthora colocasiae* (see **Fact Sheet no. 14**)

*Erwinia* species: *Erwinia carotovora* subsp. *carotovora* (see **Fact Sheet nos. 101, 214, 296**), and *Erwinia chrysanthemi*.

## Distribution

The fungi, *Athelia rolfsii* and *Lasiodiplodia theobromae* have a worldwide distribution; *Pythium splendens* and *Phytophthora colocasia* (oomycetes, water moulds, not fungi) are also recorded worldwide in tropical and sub-tropical countries. In Oceania, *Phytophthora colocasiae* is present in the Federated States of Micronesia, Guam, Northern Mariana Islands, Palau, Papua New Guinea, Samoa and Solomon Islands. *Erwinia* species also occur worldwide.

## Hosts

The fungi attack many plants. *Athelia rolfsii* attacks beans, carrots, cucumber, capsicum, peanut (see **Fact Sheet no. 11**), sweetpotato, and tomato. *Lasiodiplodia theobromae* is found worldwide, and is principally a wound pathogen, infecting underground and aboveground plant parts.

*Pythium splendens* is associated with seed rot, root rot, and seedling damping-off (see **Fact Sheet no. 47**). It is found attacking the roots of beans, cucumber, papaya, pineapple, sweetpotato, tobacco, and many more. *Phytophthora colocasia* causes a leaf blight on taro and giant taro (see **Fact Sheet no. 14**).

*Erwinia* species are bacteria with a wide host range causing soft rots. *Erwinia carotovora* pv. *carotovora* causes soft wet rots of e.g., carrots, cucurbits, Chinese cabbages (see **Fact Sheet no. 101**), lettuce (see **Fact Sheet no. 289**), potato tubers (black leg disease) (see **Fact Sheet no. 296**), tobacco, and others. *Erwinia chrysanthemi* causes soft rots in many kinds of fleshy vegetables, especially when wounded.

## Symptoms & Life Cycle

The oomycetes, *Pythium* and *Phytophthora*, infect corms through wounds made after harvest, but how this occurs is not known. Development of the rots is very rapid, and corms can be completely decayed within 4-10 days depending on their size and the number of infections.

*Pythium* rots are white, dry and crumbling, often starting at the bottom of the corm (Photo 1). There is a sharply defined, irregular boundary between the areas of rots and healthy parts. In front of the rot margin, a wide light reddish-brown area of firm healthy tissue is often present.

*Phytophthora* rots are light brown and firm (Photos 2&3)

*Athelia* rots are shallow and occur around the place where suckers have been removed, although occasionally the rots penetrate deeply into the corm. The rots show the thick white strands of the fungus growing out from the rot when corms are placed at high humidity for 24 hours (Photos 1&2).

*Lasiodiplodia* is mostly a wound pathogen associated with blights, dieback, damping-off and storage rots of all kinds of plants (Photo 4). It is often isolated from decayed corms behind the rots caused by *Pythium* and *Phytophthora*. The rots are white at first and later turn black and have a distinctive unpleasant sour smell.

*Erwinia* rots are often strong smelling white soft rots (Photo 5). They have been found in corms that were harvested and stored in leaf-lined soil pits, or in corms kept in plastic bags.

## Impact

The damage done at harvest when the "tops" and suckers are removed allows the entry of fungi, oomycetes and bacteria. The damage done by the two oomycetes is extensive and rapid; often the corms are entirely destroyed within 10 days. Invasion by *Lasiodiplodia* is also rapid, but often the fungus invades corms following colonisation by *Pythium* and *Phytophthora*. Without initial infection by *Pythium*

and *Phytophthora*, colonisation by *Lasiodiplodia* still occurs, but usually later when the corms begin to shrivel with loss of water. *Athelia* causes minor damage; the rots are much slower to develop compared to those of the other fungi and the two oomycetes. *Erwinia* rots can be very destructive, especially in corms placed at high humidity in plastic bags.

## Detection & inspection

It is difficult to detect any of the rots, except those of *Lasiodiplodia*, unless the corms are cut open. *Lasiodiplodia* can be detected as it gives the corms a sour smell, a spongy feel and, often, black powdery spore masses occur in patches on the surface of the corm.

Cut the corms in half vertically after about 7 days after harvest and storage at normal daily temperatures and humidity. Look for the white, crumbly rot of *Pythium*, the firm light brown rot of *Phytophthora*, the shallow pink rot of *Athelia*, and the black rot of *Lasiodiplodia*, usually behind rots of *Pythium* and *Phytophthora*. *Erwinia* rots are usually very characteristic: they have a foul smell, and are soft. Look for the cottony growth of *Pythium* and *Athelia* when the corms are held inside a plastic bag overnight.

## Management

### CULTURAL CONTROL

Storage life of taro corms can be extended by doing the following:

Harvest the corms leaving the "tops" and suckers attached, but remove the leaves, trimming the leaf stalks to about 10 cm. Remove large soil clods, but do not wash the corms. Place the corms in a soil pit lined with e.g., banana leaves. The pit should be situated in a shaded, well-drained location. Cover with more banana leaves. This is a traditional practice of the Sikaiana people, Solomon Islands. The corms will remain in a good condition with an acceptable taste for up to 4 weeks.

The corms stay active, and the leaves continue to grow. Presumably, leaving the "tops" and suckers in place reduces wounding and the chance of infection.

There are variations on this technique if growers want to market taro locally or overseas:

- Place corms, with "tops" and suckers attached in plastic bags. The high humidity allows the corms to grow and this prevents most fungal and oomycete rots from developing in a similar way to those in soil pits. However, *Erwinia* rots can be a problem, affecting about 10% of the corms.
- Ship the corms in plastic-lined cardboard boxes to prevent damage to the corms in transit.

Both these methods can be improved if the corms are washed and given a dip in 1% sodium hypochlorite (household bleach) for 2 minutes. A drawback of this method is that the surface colour of the corms becomes lighter than normal, which may concern some consumers. Lower concentrations of bleach or shorter treatments may be effective.

### CHEMICAL CONTROL

Other than the use of bleach, chemicals cannot be recommended for the control of storage rots; there is not enough information to show that they are cost-effective, or safe to human health. Unless plastic bags are used, the corms will always shrink and crack after treatment allowing infections of *Lasiodiplodia*. Keeping them in plastic bags is likely to be sufficient. If chemicals were to be used, checks would be needed to determine if residues were within internationally acceptable limits.

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