

Pacific Pests, Pathogens, Weeds & Pesticides - Online edition

Sugarcane smut (474)

Common Name

Sugarcane smut

Scientific Name

Sporisorium scitamineum; previously known as *Ustilago sacchari*, and *Ustilago scitaminea*. Genetic diversity appears to be greatest in Asia, but whether there are strains of the fungus is more controversial.

Distribution

Africa, Asia, North, South and Central America, the Caribbean, Europe, Oceania. It is recorded from Australia. The CABI Crop Protection Compendium notes against the record for Fiji: "Absent: Invalid presence records(s)". It was previously recorded from Fiji as *Ustilago sacchari* (Dingley JM, et al. (1981).

Hosts

Sugarcane, and related *Saccharum* species, edible and wild. While some reports mention grass species, others suggest that only *Saccharum* species are hosts.

Symptoms & Life Cycle

A serious fungal disease of sugarcane. A characteristic of the disease is the production of spore-bearing, black, whip-like structures from the growing point (Photos 1&2). Severely infected plants may be stunted with small, narrow leaves, and abundant tillers; they look weak and grass-like. More than one flush of whips can cause considerable damage to crops of susceptible varieties.

Buds either above or below ground become infected by spores (called teliospores) of the fungus. The spores germinate in water producing two types of mating spores (called sporidia); these combine to produce fungal growth which penetrates the bud. The fungus remains in the buds until cuttings are prepared, planted, and the buds begin to grow. As the buds grow, so does the fungus, and in the process it colonises the shoot. Eventually, the leaf spindle - young leaves and growing point - enclosed within the sheaths of older leaves is replaced with a long black whip, which emerges from the top of the plant. Smaller whips may develop from lateral buds giving rise to grass-like stems. At first, the sooty black-brown, roughly round spores that develop in the whip are covered by a thin membrane, but this breaks eventually to release them (Photo 3). The life-cycle is shown (Photo 4).

Spread occurs via spores blown on the wind, and from the use of infected cuttings for planting. Other possibilities are spread of spores on machinery and footwear. Survival occurs in soil for 3 to 4 months and in infected plants as long as they remain alive.

Impact

Production losses of 30 to 100% are reported depending on the susceptibility of the variety, the strain of the fungus, the environment (the disease is worse in hot dry climates) and the age of the crop when infected. There is also a loss of quality as stems may have lower sugar content. Breeding programs may be impacted because of the susceptibility of valuable lines. An approximate estimate of yield loss (in Australia) is 0.6% for every 1% increase in the number of infected plants. In the case of susceptible varieties, large numbers of dead plants can result in the loss of the ratoon crop, and additional costs due to replanting.

CABI reports that the disease appears to be cyclical, although it is not known why this is so. Epidemics of the disease are often followed by periods when the disease is of minor importance.

Detection & inspection

Look for stunted plants and the characteristic black whips that emerge from the top of the stems. ELISA and PCR-based methods are available for detection of the fungus in buds of sugarcane, and a reliable staining method using trypan blue has also given reliable results.



Photo 1. Whip-like spore-containing structure of sugarcane smut, *Sporisorium scitamineum*, emerging from the top of a plant.



Photo 2. As in Photo 1, the whip-like, spore-containing structure of sugarcane smut, *Sporisorium scitamineum*, has completely replaced the leaf spindle of the sugarcane plant.

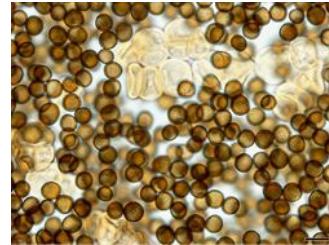


Photo 3. Spores (teliospores) of sugarcane smut, *Sporisorium scitamineum*; they are black when together on the whip, but under the microscope are brown and round.

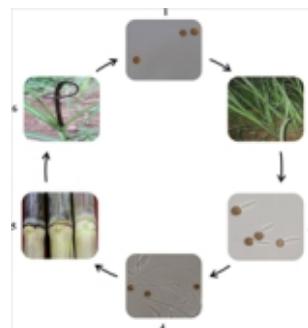


Photo 4. Infection of sugarcane smut, *Sporisorium scitamineum*. 1, Teliospores; 2, Soil and/or sugarcane plants contaminated with *Sporisorium scitamineum*; 3, Teliospore germination; 4, Germinating spores and fungal growth; 5, Infection of sugarcane meristem (growing point); 6, The formation of diseased panicle, with whip-like growth.

Management

BIOSECURITY

Because diseases of sugarcane are unevenly distributed throughout the world, all efforts should be made to prevent their further spread. Sugarcane plants moved internationally should follow the FAO/IBPGR *Technical Guidelines for the Safe Movement of Sugarcane Germplasm*. Frison EA, Putter CAJ (eds.). 1993. Rome, Italy: (http://www.bioversityinternational.org/uploads/tx_news/Musa_spp._2nd_edition_502.pdf).

CULTURAL CONTROL

Before planting

- Deep plough or irrigate fields following outbreaks to reduce the level of viable spores in the soil or in sugarcane debris. Teliospores lack dormancy, lasting only 2–3 months.
- Use hot water treatment to eliminate infections. Cuttings are immersed in water at 52°C for 30 minutes. Alternatively, 50°C for 3 hours. A method used widely for relatively small planting of sugarcane as well as large plantations. Note, however, that breeding for resistance remains the most important method for control of sugarcane smut.

During growth

- In small plantations, and at relatively low levels of infection, hot water treatment should be used in combination with removal of infected plants. Rogueing is done principally to lower infection of stems, in order to protect cuttings of the next crop.

RESISTANT VARIETIES

The production of varieties with resistance is the main method used to manage sugarcane smut. However, the level of resistance varies between locations depending on environment as dry, warm conditions favour the disease. Resistance appears to be related to the ability of spores to infect buds and/or the growth of the fungus within buds.

CHEMICAL CONTROL

This is not an appropriate method to use for controlling this disease, apart from the use of fungicides after hot water treatment. Usually, cuttings are dipped in fungicide (flutriafol or propiconazole) after hot water treatment to protect the buds, which may become soft and more easily infected.

When using a pesticide, always wear protective clothing and follow the instructions on the product label, such as dosage, timing of application, and pre-harvest interval. Recommendations will vary with the crop and system of cultivation. Expert advice on the most appropriate pesticides to use should always be sought from local agricultural authorities.

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Information from Sugar Research Australia. Sugarcane smut. Information sheet IS13012. (<https://sugarresearch.com.au/wp-content/uploads/2017/02/Sugarcane-smut-IS13012.pdf>); and CABI (2019) *Sporisorium scitamineum* (sugarcane smut). Crop Protection Compendium. (<https://www.cabi.org/cpc/datasheet/55949>); and Sugarcane smut. Wikipedia. (https://en.wikipedia.org/wiki/Sugarcane_smut); and from Dingley JM, et al. (1981) Records of fungi, bacteria, algae, and angiosperms pathogenic on plants in Cook Islands, Fiji, Kiribati, Niue, Tonga, and Western Samoa. Technical Report (2). South Pacific Bureau for Economic Co-operation, United Nations Development Programme, Food and Agricultural Organisation of the United Nations. Photo 1 William M. Brown Jr. Bugwood.org. Photos 2&3 Roger Siwas (2010) Sugarcane smut (*Ustilago scitamineae*). PaDIL - (<http://www.padil.gov.au>). Photo 4 Que Y, et al. (2014) Genome sequencing of *Sporisorium scitamineum* provides insights into the pathogenic mechanisms of sugarcane smut. BMC Genomics 2014(15):996. <https://bmcbioinformatics.biomedcentral.com/articles/10.1186/1471-2164-15-996>.

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